

## Optimization of Biochemical Treatment of Tannery Wastewater

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### Abstract

The present work is concerned with the finding of the optimum conditions for biochemical wastewater treatment for a local tannery. The water samples were taken from outline areas (the wastewater of the chrome and vegetable tannery) in equal volumes and subjected to sedimentation, biological treatment, and chemical and natural sedimentation treatment.

The Box-Wilson method of experimental design was adopted to find useful relationships between three operating variables that affect the treatment processes (temperature, aeration period and phosphate concentration) on the Biochemical Oxygen Demand (BOD5).

The experimental data collected by this method were successfully fitted to a second order polynomial mathematical model. The most favorable operating conditions for the treatment are;

Temperature	32.5oC,
Aeration period	10 hours and
phosphate concentration	16.8 mg/L.

On using the optimum conditions a mathematical model simulating the operation for the treatment was obtained.

### Introduction

In leather industry (tanning) and for practical reasons, almost all tanneries are located on rivers to provide them with process water (30 to 80 m3 for 1 ton of processed raw skins) [1] and for the disposal of their effluents. The effluent from this industry is mainly waterborne [2]. The components of the effluent arise from purification of raw hides and skins before processing as well as from residual chemicals from the production processes. Hence, the tanning industry had been recognized as a major contributor to water pollution problems.

Biodegradable organic matter consumes oxygen and nutrients in complex biochemical reactions until rendered inert [3]. This exerts a biochemical oxygen demand as one

of the fundamental parameters used to regulate the quality of the effluent [4]. The contaminants in industrial wastewater are removed by physical, chemical and biological means [5]. Facilities for handling wastewater are usually considered to have three major parts: collection, treatment and disposal [6]. Pre-aeration is used to "freshen" the wastewater and to assist the removal of oil and grease [7]. Secondary treatment processes commonly consist of biological processes. This means that living organisms which control the environment of the process are used to partially stabilize (oxidize) organic matter not previously removed by treatment processes and to convert it into a form which is easier to remove from the wastewater [8]. Sedimentation or primary treatment makes the wastewater much clearer. Two clarifiers are used to provide detention time (3 h) where, almost 60% of the suspended solids (SS) will either settle to the bottom or float to the surface and be removed. Removal of

these solids will usually reduce the BOD<sub>5</sub> of the waste approximately 35%.

The next step is the biological treatment which can typically be divided into aerobic and anaerobic. Anaerobic biological treatment is an oxygen-devoid process. Aerobic biological treatment is done in the presence of oxygen. It is applicable to wastewater containing bio-degradable organic constituents and some non-metallic inorganic constituents [2]. The activated sludge process (ASP) is the currently used biological treatment process for wastewater in Al-Za'afaraniya tanning factory, southern Baghdad. The system consists of an equalization basin, a settling tank, an aeration basin, a clarifier, and a sludge line. The recirculation of the biomass, which is an integral part of the process, allow microorganisms to adapt changes in wastewater composition with relatively short acclimation time and also allow a greater degree of control over acclimated bacterial population [10].

For a proper control of the ASP, the growth of the microorganisms should be controlled. Bacteria make up about 95% of the activated sludge biomass. These single celled organisms grow in the wastewater by consuming (eating) biodegradable materials such as proteins, carbohydrates, fats and many other compounds. Some important factors acting on growth and activity of bacteria in biochemical wastewater treatment are: food-to-microorganism ratio (F/M); use of oxygen [11]; formation of Floc [12]; mixing [12]; ; pH [13]; temperature [6] and the effect of nutrients [14]. Biological oxygen demand (BOD) is a measure of the oxygen used by microorganisms to decompose this waste. A large quantity of organic waste in the water requires large amount of bacteria to decompose it. Thus, the demand for oxygen will be high (high level of BOD). As the waste is consumed or dispersed through the water, BOD levels will begin to decline [15].

Nitrates and phosphates in a body of water can contribute to high BOD levels. Nitrates and phosphates are plant nutrients and can cause plant life and algae to grow quickly. When plants grow quickly, they also die quickly. This contributes to the organic waste in the water, which is then decomposed by bacteria. This results in a high BOD level [15]. When BOD levels are high, dissolved oxygen (DO) levels decrease because the oxygen that is available in the water is being consumed by the bacteria. Since less dissolved oxygen is available in the water, fish and other aquatic organisms may not survive [16]. The present work is an attempt to shed more light on the pollution potential of "Al-Za'afaraniya" tanning factory. The BOD<sub>5</sub> is taken as a parameter to indicate the optimum condition of biological treatment in ASP.

## EXPERIMENTAL PROCEDURE

### Materials

Phosphoric acid (70 wt%) was supplied by a local company. The chemicals used for the determination of BOD<sub>5</sub> and phosphate were of analytical grade.

An agar was used to grow bacteria, and consists of: beef extract 3 g; peptone, 5 g; agar, 15 g and distilled water to make a volume of one liter. The culture media is pH = 7.0. Malt extract agar was used to grow fungi. it consists of: malt extract, 20 g ; peptone, 5 g, agar, 15 g and distilled water to make a volume of one liter. Culture media: pH = 3.5-4.0

### Laboratory Treatment Units

Fig. 1 shows a schematic diagram of the experimental unit used for the biochemical wastewater treatment. The unit consists of: A temperature regulated water bath of (60 x 40 x 16cm) and 1.2 kW power was equipped with a stainless steel electrical stirrer (1.2 kW) and coupled with voltage regulator. The clarifier was a 3- liter Pyrex flask with a discharge valve at the bottom and a tap near the top, for the collection of treated wastewater. Air was introduced via a pre-calibrated Rotameter with and an air Sparger to ensure even distribution. A Pyrex burette was used for the delivery of nutrient (phosphoric acid). Some supplementary equipments were used including an incubator (Memmert, Germany); a microscope (Olympus); an autoclave for sterilization (Express, England); a Digital Grating Spectrophotometer (Pye unicam, England); and a Universal Pocket Meter Multiline P4 including: I. pH combined electrode with integrated temperature probe Sen tix 41; and II, Dissolved oxygen (DO) probe, Cell Ox 325.

### Procedure

The samples were taken from outline area from the wastewater of the vegetable and chrome tannery, in equal volumes. The samples were first screened to remove hair and skins pieces and then neutralized by adding sulfuric acid (50 wt%) to a final pH of (7-9). Sedimentation (settling) process was then carried out to reduce the solid content to about 65% within 2-3 h. A specified volume of the sample (30 Lit) was placed in the bath. The microorganism (activated sludge) where added to the water sample in the bath, and aeration was started.

Following the Box-Wilson method of experimental design the operating parameters used, were in the range; T=20 - 45o C  
t=5-10 hour  
PO<sub>4</sub> concentration=5-20 mg/L.

After the biochemical treatment, a polyelectrolyte at a rate of 15 mg/L was added to the mixture in the bath for a period of one hour to improve the sedimentation process of. In the natural sedimentation stage, the sample was taken from the bath and clarified for about 3 h, and then a specified volume (1.0 L) of the clarified water was taken for the BOD5 measurement.

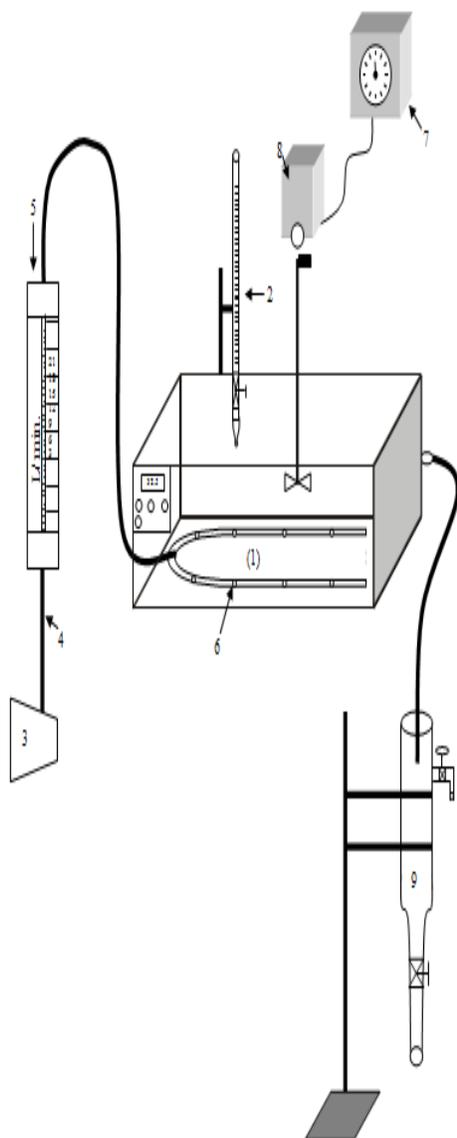


Fig. 1: The Experimental Apparatus:

1, Water bath; 2, Burette, 3, Compressor; 4, Pipes; 5, Rotameter; 6, Air sparger; 7, Voltage variator box; 8, Stirrer; 9, Clarifier

### Methods of Examination:

The TSS and volatile suspended solids (VSS) tests were done according to the WHO methods for pollution control, 1982. The (DO) concentration was measured using a (Cell DX 325) type device, which consists of a gold-metal electrode (Wissen Schuftliche Tech. Werk., Germany). The accuracy of this method is (0.1 mg/Lit). The readings were checked versus titration method with standard 0.025 N sodium thiosulfate using starch as an indicator.

The dilution, the BOD5 measurement and the determination of the phosphate concentration were carried out in accordance with standard methods [17]. The preparation of agar plate and the isolation of discrete colonies from a mixed culture were carried out according to the published methods [18].

## RESULTS AND DISCUSSION

This work deals with effect of three variables (temperature, aeration period and phosphate concentration) on BOD5.

### Postulating the Mathematical Model

A second order polynomial equation was employed in the range of the independent variables. Three variables were considered. The general form of a second polynomial equation can be given as follows:

$$Y = B_{10} + B_{11}X_{11} + B_{12}X_{12} + B_{13}X_{13} + B_{14}X_{12} X_{13} + B_{15}X_{212} + B_{16}X_{213} + B_{17}X_{11}X_{12} + B_{18}X_{11}X_{13} + B_{19}X_{12}X_{13} \quad (1)$$

The coded and real variables as well as the real values of BOD5 are given in Table 1. The data of Table 1 were fitted to equation (1) so that the regression analysis of central composite design to the approximating model to obtain the optimum conditions of the process. The coefficients of polynomial equation were evaluated. Thus, the best form of equation 1 is:

$$Y = 288.498 + 34.846 X_{11} + 18.289 X_{12} + 9.892 X_{13} + 36.896 X_{211} + 9.151X_{212} + 7.317X_{213} + 5X_{11}X_{12} + 3.75X_{11}X_{13} + 3.749X_{12}X_{13} \quad (2)$$

Correlation coefficient  $R = 0.977$ ,  
Percentage square error (S) = 1.8%.

To test the significance of each term in equation (2), the F-distribution test was used employing the variance of each term in multivariable correlation, according to Table 1 [19]. The calculations indicated insignificant interaction

between the variables (X1X3, X1X2, X2X3) are. Thus the best form of the relationship is

$$Y = 288.498 + 34.846X_{11} + 18.289X_{12} + 9.892X_{13} + 36.896X_{211} + 9.151X_{212} + 7.317X_{213} \quad (3)$$

The optimization process [20] was applied to equation (2) to find the optimum operating conditions and the results indicate the following conditions to attain the minimum BOD5 value (256 mg/L):

X11 = Temperature = 32.5o C,  
X12 = Aeration period = 10 h,  
X13 = Phosphate concentration = 16.8 mg/L.

### Effect of Different Operation Variables on the Values of BOD<sub>5</sub>:

#### Effect of Temperature:

Fig. 2 shows the influence of temperature on BOD5 at different aeration periods at fixed phosphate concentration (16.8 mg/L). It is clear that, the BOD5 decreases with increasing temperature down to a value of 250 mg/Lit at 32.5o C and aeration period of 10 h. Beyond this temperature, the BOD5 values increase. Fig. 3 shows the effects of temperature on BOD5 at various phosphate concentrations and constant aeration period of 10 h. The BOD5 reaches 250 mg/Lit at temperature of 32.5o C and phosphate concentration of 16.8 mg/L.

Fig. 4 shows the response surface function developed by the model considering temperature and aeration period. The response obtained is minimum BOD5 at temperature of 32.5o C and aeration period of 10 h. It is clear that increasing in temperature from 20o C to 32.5o C, leads to an increase in the activity of microorganisms to degrade the organic material. Beyond 32.5o C, the activity of microorganism decreases [12, 21].

#### Effect of Aeration Period:

The effects of the aeration period on the BOD5 at different temperatures and phosphate concentration are shown in Figs. 5 and 6, respectively. The BOD5 values decrease with increasing the aeration period and reaches 250 mg/L at a temperature of 32.5o C at a fixed phosphate concentration of 16.8 mg/L (Fig. 5). Fig. 6, indicates that the BOD5 reached a value of 250 mg/L at aeration period of 10 h and phosphate concentration of 16.8 mg/Lit at constant temperature of 32.5o C. The increase in the aeration period to a certain limit lowers both the BOD5 and the MLSS, since there will be longer time for the microorganism to decompose the organic matter into simpler materials [4]. However, the increase in the oxygen

supply above this limit, will improve the growth and reproduction of the microorganism and needed high cost for aeration basin [2]. The competition between these microorganism for nutrient, may lead to starvation and reduce the number of microorganism [22].

#### Effect of Phosphate:

Fig. 7 shows the influence of phosphate concentration on BOD5 at different temperatures using constant aeration period of 10 h. The BOD5 reaches a value of 250 mg/L at phosphate concentration of 16.8 mg/L and at a temperature of 32.5o C. Fig. 8 shows the influence of phosphate concentration on BOD5 at various aeration periods and constant temperature. The BOD5 decreases when phosphate concentration increases up to a level of 16.8 mg/L. Beyond this level, the BOD5 increases slowly since more phosphate concentration helps algae growth. When these algae die and decay they give organic material which increases the BOD5. Thus it may be assumed that, the input of 1 mg phosphorus lead to the growth of about 100 mg of algae dry matter [22]. Fig. 9 shows isometric relationship between aeration period, phosphate concentration with the BOD5. The response obtained is minimum (BOD5) at aeration period 10 h and phosphate concentration of 16.8 mg/L. Microorganisms require certain nutrients for growth. The basic nutrients of abundance in normal raw sewage are carbon (C), nitrogen (N) and phosphate (P) with the ratio of C:N:P of approximately 100:10:1 [23]. Phosphorus is an essential element, it is part of the structure of DNA and RNA, and is an important intermediate in metabolism [24].

Bacteriological tests indicated the presence of bacteria and protozoa in the ASP. The types of bacteria found on the surface after incubation were:

I. Staphylo cocci spp. and II. Bacilli. Meanwhile, fungi did not appear on culture, since they favour pH lower than the pH of ASP.

Applications of Optimum Condition for Wastewater Treatment Plant Plant:

After carrying out the preliminary processes, the wastewater samples were taken from the effluent of two tanneries, with their BOD5 values reduced from 760 mg/L to 610 mg/L. The optimum conditions that have been obtained above

(Temperature = 32.5o C; Aeration period = 10 h; and Phosphate concentration = 16.8 mg/L)

were used for the biochemical treatment. The treatment resulted in a decrease in the BOD5 from 436 mg/Lit to 75 mg/L in laboratory. However, the actual BOD5 value of the effluent from the first aeration basin in plant was 100 mg/L.

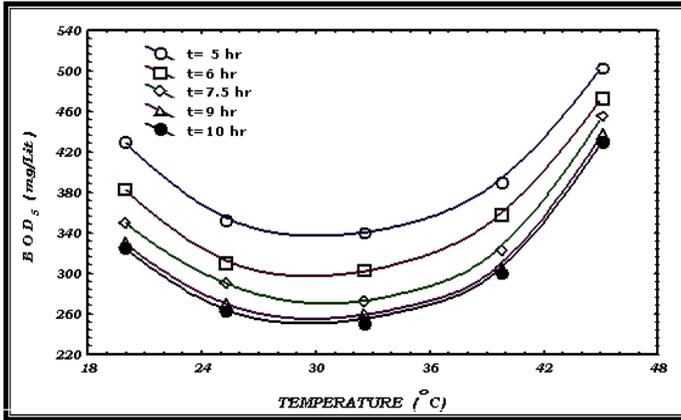


Fig.2: Effect of temperature on BOD5 at constant phosphate level Concentration (16.8mg/L)

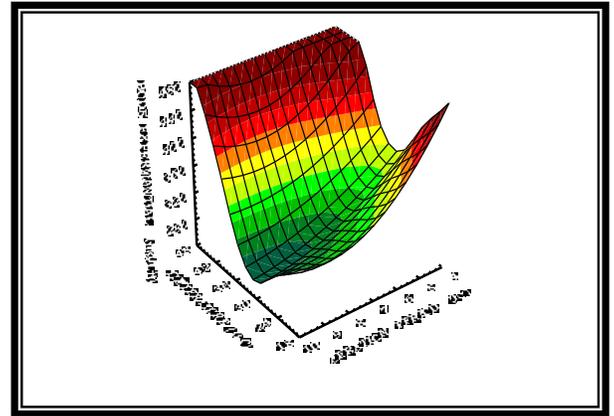


Fig.4: Effect of Temperature and aeration period on BOD5.

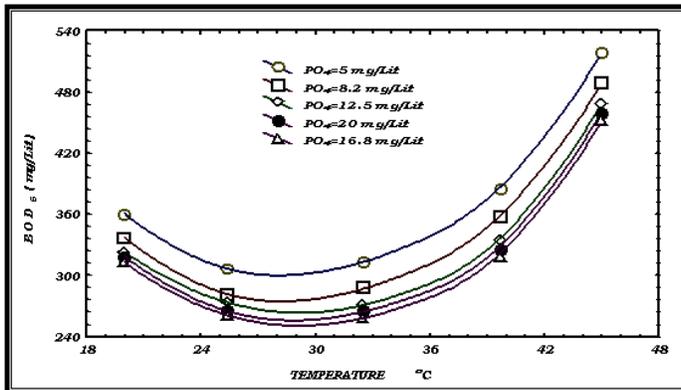


Fig.3: Effect of temperature on BOD5 at constant aeration period (10 h).

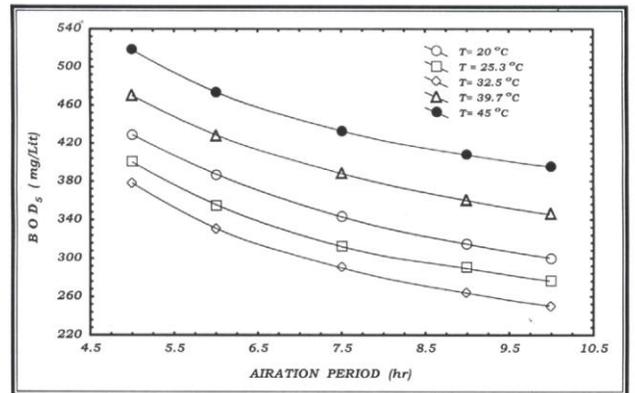


Fig.5: Effect of aeration period on BOD5 at constant phosphate concentration (16.8 mg/L)

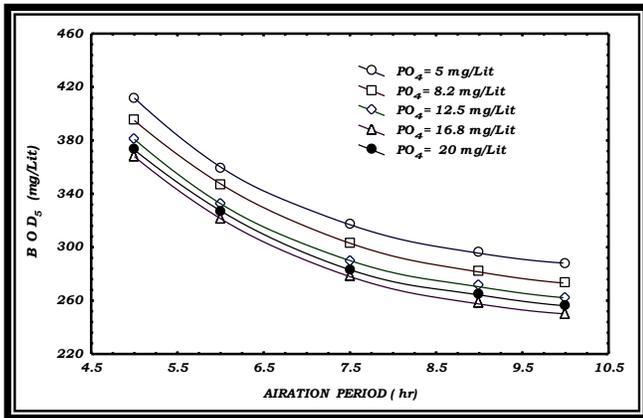


Fig. 6: Effect of aeration period on BOD5 at constant temperature (32.5 °C)

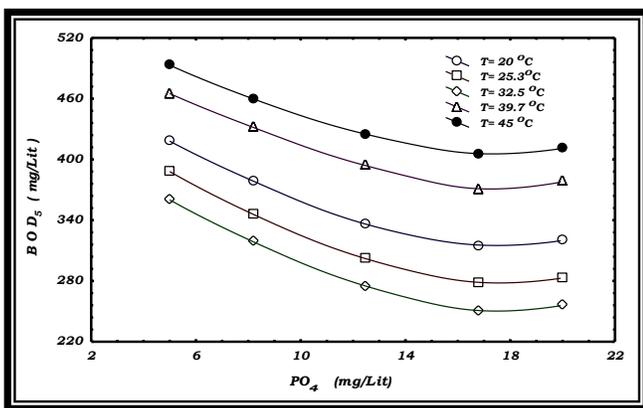


Fig.7: Effect of phosphate concentration on BOD5 at constant aeration period (10 h)

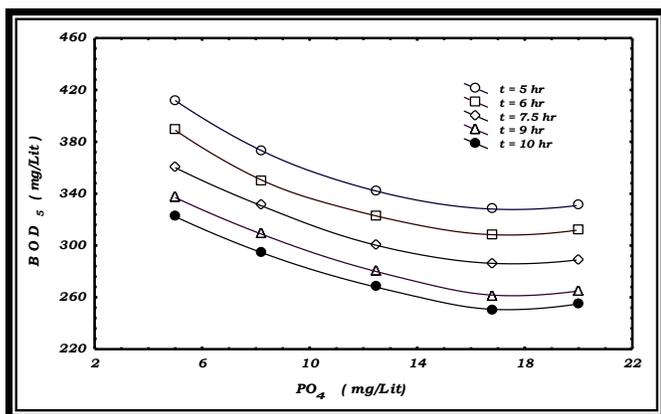
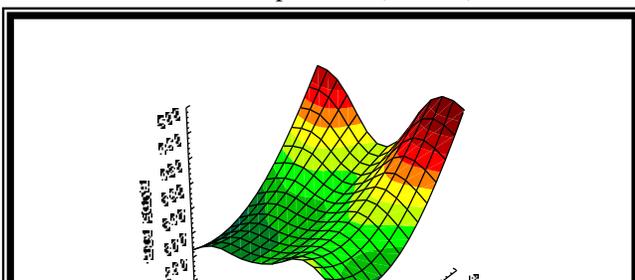


Fig. 8: Effect of phosphate concentration on BOD5 at constant temperature (32.5 °C)



## CONCLUSIONS

1. The optimum conditions for biochemical wastewater treatment the case under study were: Temperature, 32.5° C; Aeration period 10 hr and Phosphate concentration, 16.8 mg/L.
2. The biochemical oxygen demand value could be reduced from 436 mg/L down to a value of 30 mg/L after biochemical treatment at the application of optimum conditions.
3. Only bacteria and protozoa were found as microorganisms in activated sludge, because the alkaline condition and unaeration are not suitable for the living of fungi.

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